

UNIVERSITY OF THE PACIFIC
Environmental Engineering Research Program

**San Joaquin River
Up-Stream DO TMDL Project**

Task 4
Monitoring and Associated Studies
Interim Report No. 1
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Introduction

For many years, the Deep Water Ship Channel (DWSC) on the San Joaquin River (SJR) has had intermittent low dissolved oxygen (DO) conditions. The DO sag is most acute during the late summer and early fall, but low DO incidences occur year-round (Foe et al. 2002, Lehman 2001). The low DO conditions are impacting critical fish habitat and a total maximum daily load (TMDL) Implementation Plan for oxygen-demanding substances is currently being developed by the Central Valley Regional Water Quality Control Board (CVRWQCB).

In support of the development of a scientific DO TMDL allocation, 13 research and monitoring projects examining various aspects of DO demand in the SJR were conducted in the summers of 1999, 2000, and 2001. Additionally, the CVRWQCB generated a “strawman” allocation of responsibility report (Strawman Report). The Strawman Report represents a process by which responsibility for the low DO conditions can be assigned and a plan for remediation of the DO sag in the SJR can be implemented (Foe et al. 2002). The final reports for these projects can be found on the SJR DO TMDL website (www.sjrtdotmdl.org).

Studies conducted in the summers of 1999, 2000 and 2001 identified four major factors contributing the formation of a DO sag in the DWSC: the deepening of the ship channel, ammonia discharges from the Stockton Wastewater Treatment Plant (WWTP), transport of oxygen-consuming materials from the upper SJR into the DWSC, and production of oxygen-consuming organic matter in the channel. The actual impact of these factors is dependent on flow and temperature, where lower flows and higher temperatures allow a DO deficit to accumulate if the other factors are present.

The Up-Stream DO TMDL Project (ERP-02D-P63) is an inter-organizational research program focused on understanding the sources of oxygen-consuming materials in the SJR upstream of the DWSC. The purpose of this study is to provide a comprehensive understanding of the sources and fate of oxygen-consuming materials, particularly algae biomass, in the SJR watershed between Channel Point (lat long) and Lander Avenue (lat long). The study objectives are to understand current (baseline) conditions of the basin, determine the sources and sinks of oxygen demanding materials, and provide scientific tools for measuring the impact of water quality management programs being implemented in the SJR basin.

Previous studies have identified algal biomass as the most significant oxygen-demanding substance in the SJR upstream of the DWSC (Kratzer et al. 2004, Lehman 2001). Algal biomass is not a conserved substance, but grows and decays in the SJR; hence, characterization of oxygen-demanding substances in the SJR is inherently complicated and requires an integrated effort of extensive monitoring, scientific study, and modeling. The Up-Stream DO TMDL Project includes a coherent and comprehensive study of algal growth dynamics in the SJR and will identify sources of algal nutrients to the SJR.

Other oxygen-demanding substances found in the SJR above the DWSC include ammonia and organic carbon from sources other than algae (Kratzer et al. 2004). The SJR watershed contains municipalities, dairies, wetlands, and agricultural industries that may potentially contribute biochemical oxygen demand (BOD) to the SJR. This study is designed to discriminate between algal BOD and other sources of BOD throughout the entire SJR watershed.

This interim report concerns activities conducted under Task 4 (Monitoring and Scientific Studies) of the Up-Stream DO TMDL Project. The specific objectives of Task 4 are to:

- 1) Collect sufficient hydrologic and water quality data to characterize the loading of algae, other oxygen-demanding materials, and nutrients from individual tributaries and sub-watersheds of the upstream SJR between Channel Point and Lander Avenue.
- 2) Identify and characterize sub-watersheds that are the most significant sources of algal biomass, nutrients, and BOD to the river, providing basic scientific information to support a water quality management program.
- 3) Characterize the transformation and fate of nutrients and algae in the watershed and the impact of nutrients on algal growth.
- 4) Characterize the temporal variability of water quality parameters on a daily and seasonal basis.
- 5) The data collected in Task 4 will also be used in Task 6 to model and predict the transformation and fate of algae in the river.

Scientific activities for Task 4 commenced in the early spring of 2005 and this project is currently in seventh month of research. The initial focus of Task 4 was the development and execution of a bi-monthly (twice-monthly) grab sampling program for the collection of water-quality data. This program began in March 2005 and this report includes a description of that program and water-quality data collected between March and July 2005. Research to characterize individual sub-watersheds has been started. This research is in its preliminary stages and results from those efforts are not included in this report, but will be described in the second Interim Report scheduled for March 2006. In cooperation with activities conducted under Task 5 (Installation of Flow Monitoring Stations), scientists under Task 4 have been compiling and evaluating flow and electrical conductivity data generated by watershed groups. That work under Task 4 has consisted largely of compiling raw data and training technical personnel on station calibration and quality control procedures. Scientific effort on characterization of temporal and seasonal variability are initiated, but results are too premature to warrant inclusion in this report. A characterization of algal and nutrient interactions premature until more data is collected.

Methods

Grab samples and field measurements were executed in accordance with guidelines from the SWRCB (Puckett 2002). Grab samples from each site are collected in a minimum of 3, one liter glass, acid washed Wheaton bottles, as well as a minimum of 3 250 mL plastic bottles (VWR trace-clean). The 1 liter Wheaton bottles are filled by attaching one open bottle to a custom-built cradle attached to a 14 foot telescoping pole. The bottle is then slowly dipped into the site waters to effect the collection of a depth-integrated sample. This process is then repeated for the remaining 1 L Wheaton bottles. The 250 mL bottle samples are collected via a custom-built cradle attached to a similar 14 foot telescoping pole. All three bottles are held in the cradle contemporaneously and dipped into the site waters to effect the collection of a depth-integrated sample. For sampling sites inaccessible with the telescoping pole, a 2 L bucket attached to a rope is dropped into the site waters and retrieved. The bucket's contents are then poured into the sample bottles by swirling the water in the bucket while pouring into a funnel that is randomly rotated from sample bottle to sample bottle. This alternative process is used to fill both the 1 L Wheaton and the 250 mL bottles. All sample bottles are capped immediately after a sample has

been collected, placed in an ice cooler, and kept at a temperature of 4° Celsius (C) until delivered to the laboratory.

Light is measured using both a VWR LUX light-meter and a custom-built PAR platform incorporating a Li-Cor spherical quantum detector. The LUX light-meter measures light in the visible spectrum the way a photographer would for exposing film. The PAR platform measures sunlight as photosynthetically-active radiation in micro-Moles per second per square meter of photons. Each device is placed in open ground, under direct sunlight, in as flat a position as possible. The PAR meter has an incorporated bubble site level to better achieve this latter criterion.

Sample site velocity is recorded using a Marsh-McBirney Flo-mate 2000 (measured in feet/second). The sensor head of this device is attached in a perpendicular position to the end of a six-foot pole, which is then dipped into the site waters to a depth of 40% total stream depth or at least two feet from the surface in deeper water. The sensor head, with the help of a visual guide, is then positioned to face upstream and collect data.

A multi-parameter YSI 6600 SONDE is used to collect temperature (°C), specific conductivity (mS/cm), total dissolved solids (g/L), dissolved oxygen (sat %, mg/L, charge), pH, depth (feet), oxidation-reduction potential (mV), turbidity (NTU), chlorophyll-a (ug/L), Fluorescence (%FS), and barometric pressure (mm HG) data. The sonde is held vertically in the site waters, deep enough to submerge the device, but shallow enough to prevent it from resting on the bottom. The data is then collected via a cable connection to a YSI MDS-650 handset and recorded internally. Each sonde undergoes a pre- and post-operation calibration in the laboratory to ensure consistent and accurate data collection.

All samples were processed and/or preserved within 24 hours of sampling. Processing consisted of filtering the samples using a vacuum filter apparatus with a Whatman GF/F filter, preservation of TOC samples, and setting up the BOD analyses. Loading of sample onto the filter was dependent on the amount of solids in the sample and ranged from 250 to 1000 ml of water per filtration. Filters were prepared for chlorophyll analysis, USGS analysis, and TSS/VSS determination. Filtrate was collected for DOC and USGS analysis, as well as for internal archive. For all analyses, standard curves and other quality control parameters were followed as outlined in the QAPP for this project.

Biochemical oxygen demand (BOD) was analyzed by Standard Method (SM) 52101 B with a modification for measurement of oxygen demand at 10 days rather than 5 days (APHA 1998). Previous studies in the SJR have used 10-day BOD analysis (BOD_{10}) as a standard procedure and it is necessary to continue this protocol order to be consistent with prior studies. The BOD_{10} tests were initiated within 24 hours of sampling. Samples were kept at 4°C until incubation, but were stabilized at 20°C before the initial oxygen reading. The samples were shaken prior to incubation to assure initial dissolved oxygen (DO) levels between 7 and 9 mg/L. BOD_{10} was measured on all samples without seed, also as in previous studies. Carbonaceous BOD ($CBOD_{10}$), was determined by inhibiting nitrogenous demand with N-Serve Nitrification Inhibitor (HACH, Loveland, Colorado). Nitrogenous BOD ($NBOD_{10}$) was then determined by subtracting $CBOD_{10}$ from BOD_{10} . Initial and final dissolved oxygen was measured using a calibrated YSI 5100 DO meter (Yellow Springs, Ohio). One duplicate measurement and three blank measurements (trip blank, buffer solution, and deionized water) were prepared for a minimum of every sample set or every 20 samples. Samples were run both full strength and diluted 1:3 to assure that a valid BOD_{10} measurement was achieved for all samples, with a

residual greater DO than 1 mg/L and at least 1 mg/L consumed. Standards were prepared with each standard set consisting of buffer solution spiked with a known amount of 300mg/l glucose/300mg/l glutamic acid (HACH, Loveland, CO) and 5 ml sample seed.

Total organic carbon (TOC) was measured by high temperature combustion using a Teledyne Tekmar Apollo 9000 (Mason, Ohio) according to SM 5310 B (APHA 1998). Dissolved organic concentration (DOC) phosphoric acid and stored in the dark at 4°C. Deionized water rinses were run between all samples to reduce occurrence of sample carryover. Samples were run within 28 days of collection and preservation.

Total suspended solids (TSS) and volatile suspended solids (VSS) were analyzed by SM 2540 D and E, respectively (APHA 1998). The filters were pre-rinsed with deionized water, placed in aluminum dishes, and pre-combusted at 550°C for 1 hour prior to use. After combustion the filters and aluminum dishes were stored in a dessicator until use. Sample loading was dependent on the amount of solids in the sample and generally ranged from 250 to 1000 ml of sample per filter. After filtration, samples were dried in a vacuum oven at 105°C until a constant weight was achieved, then combusted at 550°C for 1 hour.

Chlorophyll-a (chl-a) and pheophytin-a (pha-a) was extracted and analyzed using UV absorption (SM 10200H). Filtration of the samples was done under low light conditions and sample loaded onto the filter was preserved with a saturated MgCO₃ solution and storing at -20°C. Filters were ground in a 90% acetone/10% MgCO₃ solution using a Teflon tissue grinder while maintaining the samples below 4°C at all times. The ground filters were extracted in the acetone mixture at 4°C for 4 to 24 hours. The extract was analyzed on a Perkin Elmer Lambda 33 Spectrophotometer (Boston, MA). After measurement of the extract, 33microliters of 0.1N HCl/ml of sample were added for determination of pheophytin. Analysis of sample was performed within 14 days of sampling.

Nitrate and ammonium concentrations are determined on samples filtered through a 0.45 mm Nuclepore membrane filter (filters are pre-rinsed with sample). Nitrate and ammonium are quantified simultaneously using an automated membrane diffusion/conductivity detection method (Carlson, 1978, 1986; Carlson et al., 1990).

Total nitrogen was determined on non-filtered samples. Total nitrogen is determined conductimetrically (as described above) following persulfate oxidation (Yu et al., 1994) using a 1% persulfate oxidant concentration, a sample:oxidant ratio of 1:1 (V/V), and heating in an autoclave. Recovery of total nitrogen is statistical identical to the Kjeldahl total nitrogen method in a comparison study conducted by UC Davis utilizing several reagent grade, organic nitrogen compounds. Quantification of nitrogen is by the automated membrane diffusion/conductivity detection method (Carlson, 1978, 1986; Carlson et al., 1990).

Ortho-phosphate is determined on samples filtered through a 0.45 mm Nuclepore membrane filter (filters are pre-rinsed with sample). We use method SM 4500-P.D, the “stannous chloride” method for this analysis (APHA 1998). We have the ability to utilize a 5 cm cell to lower the limit of detection; however, we find that the 1 cm cell is sufficient for most Central Valley river waters. Alternatively, ortho-phosphate and total phosphorous will be quantified by the Ascorbic Acid Method (adapted from SM 4500-P-E).

Total phosphorus is determined on non-filtered samples. Total P is measured by the “stannous chloride” method following persulfate digestion as described above for the total N procedure. The limit of detection for this method is about 5 ppb P using a 1 cm cell for measurement.

Results

A major effort of Task 4 has been the collection of water quality data from 20 sites that are believed to represent the San Joaquin River and its major tributaries. The location of these twenty “core” grab sampling sites and a brief description are provided in Table 1. An overall map of the Core Sites is provided in Figure 1. In Figures 2 through 20, aerial pictures of the site locations are shown. In addition to these Core Sites, grab samples were collected at Fremont Ford on the SJR and Modesto Irrigation District Lateral 4 during the period included in this report (March to July 2005). Water quality results for these sites are summarized in Table 2. Photographs documenting some of the sampling activities conducted under Task 4 are shown in Figures 21 through 27.

Station installation activities have begun under Task 5 and raw data has been provided to the Task 4 team by water districts on the Westside of the SJR. East-side flow stations are being up-graded and stage and electrical conductivity (EC) will become available in the next few months. Flow and EC data from stations managed by the USGS and DWR will be compiled as it becomes available in the fall. Table 3 provides a summary of flow stations identified for potential inclusion in this study.

An example of some of the raw data being provided by the new West-side stations is shown in Figure 28 and 29. Calculation of flow for each station is dependent on the calibration of the stage-flow relationship, which is in progress, so quality assured flow data can not be provided in this report. A quality control program for continuous EC measurement is being implemented and raw EC data will be processed using statistical methods developed for other projects.

Discussion

The SJR basin is located in a Mediterranean climate characterized by a dry season (typically May through November) and a wet season (November through May). During the dry period, the water quality in the river is heavily influenced by return flows from irrigated lands. With the exception of the Tuolumne, Stanislaus, and Merced Rivers, which convey water from the Sierra reservoirs, the tributaries included in this study consist largely of irrigated runoff during the dry season.

The results collected between March and July largely support previous studies that indicate algal biomass, as measured by chlorophyll-a (chl-a), is a major contributing factor to BOD in the basin ($r^2 = 0.523$), however the poor relationship between total organic carbon (TOC) and chl-a ($r^2 = 0.229$) and the strong relationship between BOD and TOC ($r^2 = 0.513$) suggest that other sources may also contribute to BOD in the river. Results from individual tributaries (Table 2) also indicate that algae are not the only source of BOD in the system. The relationship between BOD, TOC, and chl-a is being further investigated.

Results presented in Table 2 indicate that variability in water quality within a tributary is low enough that significant and consistent differences between tributaries can be measured with a bi-monthly grab sampling program. These results suggest that the concept of developing a predictive model for the basin has a significant probability for success. Future research will investigate inter-year variability and differences between wet and dry season water quality.

Further investigation of additional tributaries will be designed to allow statistical comparison to the Core Sites, with the intention of determining if water quality results from the Core Sites can be used to characterize tributaries that are sampled less frequently. Currently, models of the SJR are using data from a few sites to estimate water quality at uncharacterized tributaries without clear evidence that this approach is valid. The validity of this approach should be resolved as the result of research carried out under Task 4.

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Table 1: Location and common name of the Core Sample Sites for the DO TMDL Project. These sites are scheduled for bi-monthly summer sampling for the duration of the project. Sample locations that were also included in prior water quality studies (Kratzer et al. 2004) are indicated in the right column.

DO Site Number	Sample Station Name	Latitude	Longitude	Included in UC Davis & USGS Study
4	SJR at Mossdale	37.7871	-121.3076	X
5	SJR at Vernalis	37.6758	-121.2653	X
6	SJR at Maze	37.6400	-121.2292	X
7	SJR at Patterson	37.4937	-121.0808	X
8	SJR at Crows Landing	37.4320	-121.0117	X
10	SJR at Lander Avenue	37.2942	-120.8513	
12	Stanislaus River at Caswell Park	37.7016	-121.1772	X
14	Tuolumne River at Shiloh Bridge	37.6035	-121.1313	X
16	Merced River at River Road	37.3504	-120.9620	X
18	Mud Slough near Gustine	37.2625	-120.9056	X
19	Salt Slough at Lander Avenue	37.2480	-120.8519	
20	Los Banos Creek at Highway 140	37.2763	-120.9557	X
21	Orestimba Creek at River Road	37.4140	-121.0149	X
23	Modesto ID Lateral 5	37.6145	-121.1434	
25	Modesto ID Miller Lake	37.6702	-121.2193	
28	Turlock ID Westport Drain	37.5420	-121.0941	
29	Turlock ID Harding Drain	37.4643	-121.0309	
36	Del Puerto Creek Flow Station	37.5395	-121.1221	
44	San Luis Drain End	37.2609	-120.9052	X
59	SJR Laird Park	37.5573	-121.1501	X

Table 3: Current information on flow measurements and flow data availability for the San Joaquin River watershed. Table includes flow & grab sample stations originally included in the March 2003 proposal as well as additional stations or tributaries identified in field surveys and meetings with watershed groups conducted between February and July, 2005. Stations designated “NA” have not yet been assigned site numbers for the DO TMDL Project.

DO Site Number	Sample Station Name	Latitude	Longitude	Organization Responsible for Flow Station	Suggested Contact for Flow Data
1	SJR at Channel Point	37.95027	-121.33715	No station	None
2	SJR at Lathrop	37.86488	-121.32267	DWR	Joe Tapia
3	SJR at Old River	37.81082	-121.32392	DWR	Joe Tapia
4	SJR at Mossdale	37.78710	-121.30757	DWR	Joe Tapia
5	SJR at Vernalis-McCune Station (River Club)	37.67936	-121.26504	DWR (use Vernalis)	USGS Sacramento
6	SJR at Maze	37.64142	-121.22902	USGS	USGS Sacramento
7	SJR at Patterson	37.49373	-121.08081	DWR	Joe Tapia
8	SJR at Crows Landing	37.43197	-121.01165	USGS	SFEI
9	SJR at Fremont Ford	37.30985	-120.93055	DWR	SFEI
10	SJR at Lander Avenue	37.29424	-120.85125	DWR	Joe Tapia
11	French Camp Slough-Van Buskirk	37.91613	-121.30447	No Station	None
12	Stanislaus River at Caswell Park	37.70160	-121.17719	No Station (use Ripon)	USGS Sacramento
13	Stanislaus River at Ripon	37.73113	-121.10811	USGS	USGS Sacramento
14	Tuolumne River at Shiloh Bridge	37.60350	-121.13125	No Station (use Modesto)	USGS Sacramento
15	Tuolumne River at Modesto	37.62722	-120.98742	USGS	USGS Sacramento
16	Merced River at River Road	37.35043	-120.96196	No Station (use Stevinson)	DWR/Joe Tapia
17	Merced River near Stevinson	37.38730	-120.79366	DWR	Joe Tapia
18	Mud Slough near Gustine	37.26250	-120.90555	USGS	SFEI
19	Salt Slough at Lander Avenue	37.24795	-120.85194	USGS	SFEI
20	Los Banos Creek Flow Station	37.27546	-120.95532	Grassland WD	Lara Sparks
21	Orestimba Creek at River Road	37.41396	-121.01488	Del Puerto WD	Liz Vonckx
22	Modesto ID Lateral 4	37.63057	-121.15888	Modesto ID	Michael Niemi
23	Modesto ID Lateral 5	37.61452	-121.14339	Modesto ID	Michael Niemi

Table 3: (continued) Flow measurement information.

DO Site Number	Sample Station Name	Latitude	Longitude	Organization Responsible for Flow Station	Suggested Contact for Flow Data
24	Modesto ID Lateral 6 to Stanislaus River	37.70383	-121.14143	Modesto ID	Michael Niemi
25	Modesto ID Miller Lake	37.66792	-121.21520	Modesto ID	Michael Niemi
26	Turlock ID Highline Spill	37.38921	-120.80568	Turlock ID	Keith Larson
27	Turlock ID Lateral 2	37.56522	-121.13836	Turlock ID	Keith Larson
28	Turlock ID Westport Drain	37.54196	-121.09408	No Station	TID/Keith Larson
29	Turlock ID Harding Drain	37.46427	-121.03093	Turlock ID	Keith Larson
30	Turlock ID Lateral 6 & 7 at Levee	37.39782	-120.97225	No Station	TID/Keith Larson
31	BCID – New Jerusalem Drain	37.72669	-121.29963	SJVDA	Liz Vonckx
32	El Solyo WD – Grayson Drain	37.58563	-121.17699	No Station	None
33	Hospital Creek	37.61029	-121.23082	SJVDA	Liz Vonckx
34	Ingram Creek	37.60026	-121.22506	SJVDA	Liz Vonckx
35	Westley Wasteway Flow Station	37.55818	-121.16375	SJVDA	Liz Vonckx
36	Del Puerto Creek Flow Station	37.53947	-121.12206	SJVDA	Liz Vonckx
37	Newman Wasteway	37.33768	-120.97207	No Station	None
38	Marshall Road Drain	37.43605	-121.03600	SJVDA	Liz Vonckx
39	Salado Creek at River Rd	37.49960	-121.10539	No Station	None
40	Patterson Irrigation District Diversion	37.49716	-121.08280	PID	John Sweigart
41	West Stanislaus Irrigation District Diversion	37.58438	-121.20053	WSID	Ron Roos
42	Banta Carbona Irrigation District Diversion	37.71266	-121.31146	BCID	David Wisenberger
43	El Solyo Water District Diversion	37.64011	-121.22949	ESWD	John Hanson (DPWD)
44	San Luis Drain End	37.26090	-120.90520	No Station (Use Station B)	SFEI
45	Volta Wasteway at Ingomar Grade	37.10528	-120.93643	Grassland WD	Lara Sparks
45	Volta Wasteway Flow Station	37.12903	-120.91937	Grassland WD	Lara Sparks

Table 3: (continued) Flow measurement information.

DO Site Number	Sample Station Name	Latitude	Longitude	Organization Responsible for Flow Station	Suggested Contact for Flow Data
46	Mud Slough at Gun Club Road	37.23145	-120.89923	Grassland WD	Lara Sparks
47	Delta-Mendota Canal inlet to the Mendota Pool	36.78070	-120.37221	Exchange Contractors Authority	Nigel Quinn
48	FC-5 – Grassland Area Farmers	36.92428	-120.65411	SJVDA	Joe McGahan, Mike Gardener
49	PE-14 – Grasslands Area Farmers	36.93884	-120.63555	SJVDA	Joe McGahan, Mike Gardener
50	San Luis Drain Site A	36.96660	-120.67060	GBP	SFEI
51	Arroyo Canal	37.08526	-120.81582	Exchange Contractors Authority	Nigel Quinn
52	Salt Slough at Sand Dam	37.12415	-120.73735	No Station	None
53	Salt Slough at Wolfsen Road	37.15937	-120.81292	LBNL	Nigel Quinn
54	Los Banos Creek at Ingomar Grade	37.07780	-120.88046	No Station	None
55	Modesto WWTP	37.53584	-121.09311	City of Modesto	NPDES
56	Turlock WWTP	37.48421	-120.87039	City of Turlock	NPDES
59	SJR Laird Park	37.55731	-121.15011	No Station	None
NA	DP-25 – Grasslands Area Farmers	36.86626	-120.63584	SJVDA	Joe McGahan, Mike Gardener
NA	San Luis Drain Site B	37.24082	-120.88190	USGS	SFEI
NA	Tartar Plant	37.32000	-120.98000	Unknown	NPDES
NA	Newman WWTP	37.32304	-120.98233	City of Newman	NPDES
NA	SJR Hills Ferry	37.34944	-120.97520	USGS	USGS Sacramento
NA	Turlock ID Stevinsen Spill Flow Station	37.37129	-120.93070	Turlock ID	Keith Larson
NA	Moran Drain	37.43547	-121.03551	SJVDA	Liz Vonckx
NA	Spanish Grant Drain	37.43576	-121.03581	SJVDA	Liz Vonckx
NA	Ramona Lake	37.47881	-121.06850	SJVDA	Liz Vonckx
NA	Patterson WWTP	37.49989	-121.09140	City of Patterson	NPDES

Table 3: (continued) Flow measurement information.

DO Site Number	Sample Station Name	Latitude	Longitude	Organization Responsible for Flow Station	Suggested Contact for Flow Data
NA	Modesto ID Jacobsen Drain	37.61657	-121.15910	Modesto ID	Michael Niemi
NA	El Solyo Water District Drain	37.64060	-121.22925	No station	None
NA	Modesto ID Main Drain to Miller Lake	37.66619	-121.19553	Modesto ID	Michael Niemi
NA	SJR at Vernalis (Flow Station)	37.67578	-121.26527	USGS	USGS Sacramento
NA	Banta WWTP	37.75313	-121.32589	City of Banta	NPDES
NA	Manteca WWTP	37.79540	-121.30905	City of Manteca	NPDES
NA	Manteca Storm Drain	37.80308	-121.31260	City of Manteca	NPDES
NA	Old River USGS Flow Station	37.80782	-121.33149	USGS	USGS Sacramento
NA	French Camp Slough-Airport Way	37.88154	-121.24914	DWR	Joe Tapia
NA	SJR Brickyard Site	37.89723	-121.32736	University of Pacific	Gary Litton
NA	SJR at Stockton (Garwood Bridge)	37.93495	-121.32940	USGS	USGS Sacramento
NA	Stockton WWTP	37.93810	-121.33580	City of Stockton	NPDES
NA	SJR at Rough and Ready Island	37.96257	-121.36600	DWR	Joe Tapia

Figure 1: Map of DO TMDL Project Core Sites for water quality sampling. See Table 1 for site names.

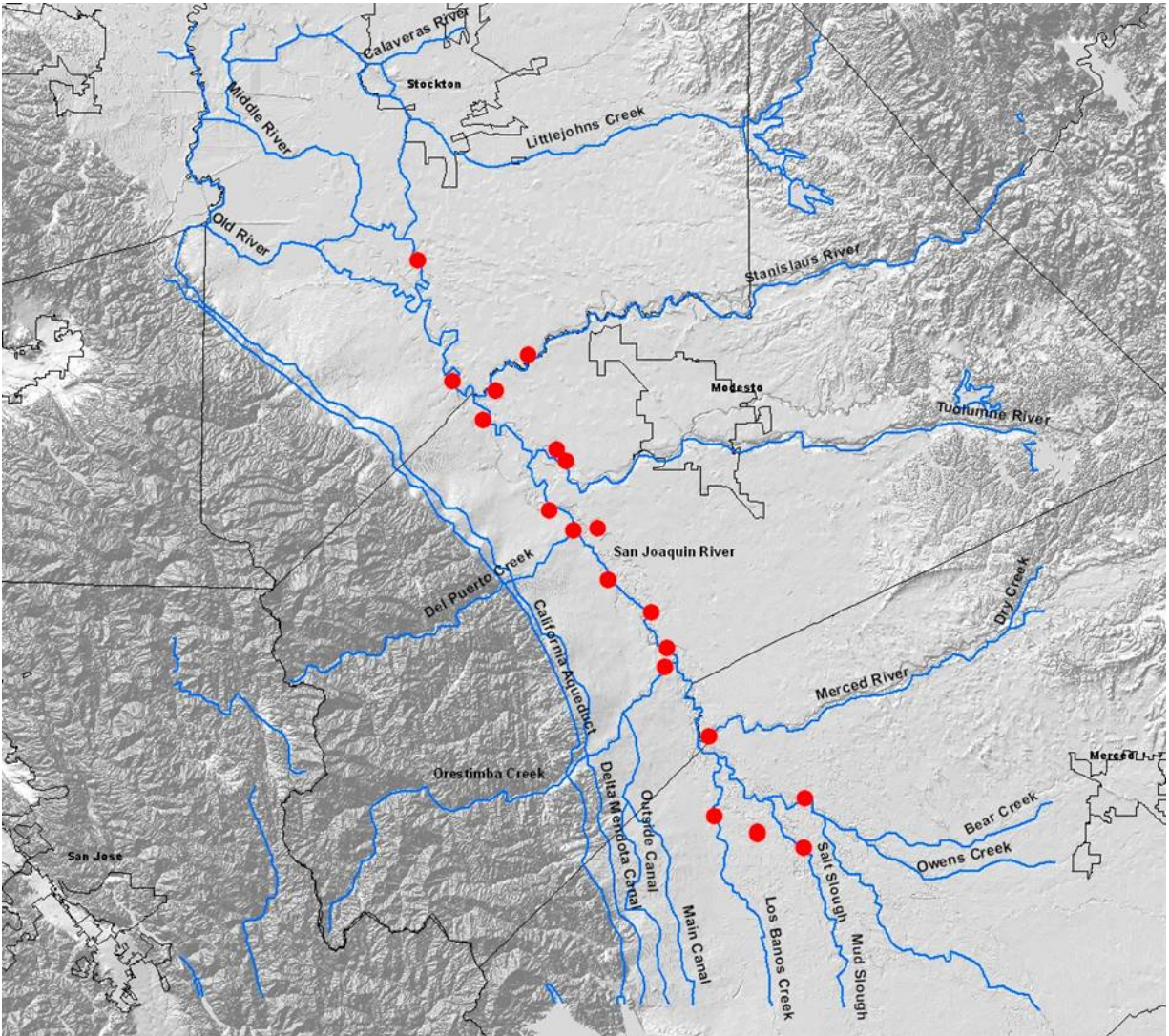


Figure 2: Aerial view of DO TMDL Project water quality sampling station DO-4 San Joaquin River at Mossdale.



Figure 3: Aerial view of DO TMDL Project water quality sampling station DO-5 San Joaquin River at Vernalis.



Figure 4: Aerial view of DO TMDL Project water quality sampling station DO-6 San Joaquin River at Maze Boulevard.



Figure 5: Aerial view of DO TMDL Project water quality sampling station DO-7 San Joaquin River at Patterson.



Figure 6: Aerial view of DO TMDL Project water quality sampling station DO-8 San Joaquin River at Crows Landing.



Figure 7: Aerial view of DO TMDL Project water quality sampling station DO-10 San Joaquin River at Lander Avenue.



Figure 8: Aerial view of DO TMDL Project water quality sampling station DO-12 Stanislaus River at Caswell Park.



Figure 9: Aerial view of DO TMDL Project water quality sampling station DO-14 Tuolumne river at Shiloh Bridge.



Figure 10: Aerial view of DO TMDL Project water quality sampling station DO-16 Merced River at River Road.



Figure 11: Aerial view of DO TMDL Project water quality sampling station DO-18 Mud Slough near Gustine and DO-44 San Luis Drain End.



Figure 12: Aerial view of DO TMDL Project water quality sampling station DO-19 Salt Slough at Lander Avenue.



Figure 13: Aerial view of DO TMDL Project water quality sampling station DO-20 Los Banos Creek at Highway 140.



Figure 14: Aerial view of DO TMDL Project water quality sampling station DO-21 Orestimba Creek at River Road



Figure 15: Aerial view of DO TMDL Project water quality sampling station DO-23 Modesto Irrigation District Lateral 5 at Paradise Road.



Figure 16: Aerial view of DO TMDL Project water quality sampling station DO-25 Modesto Irrigation District Miller Lake.

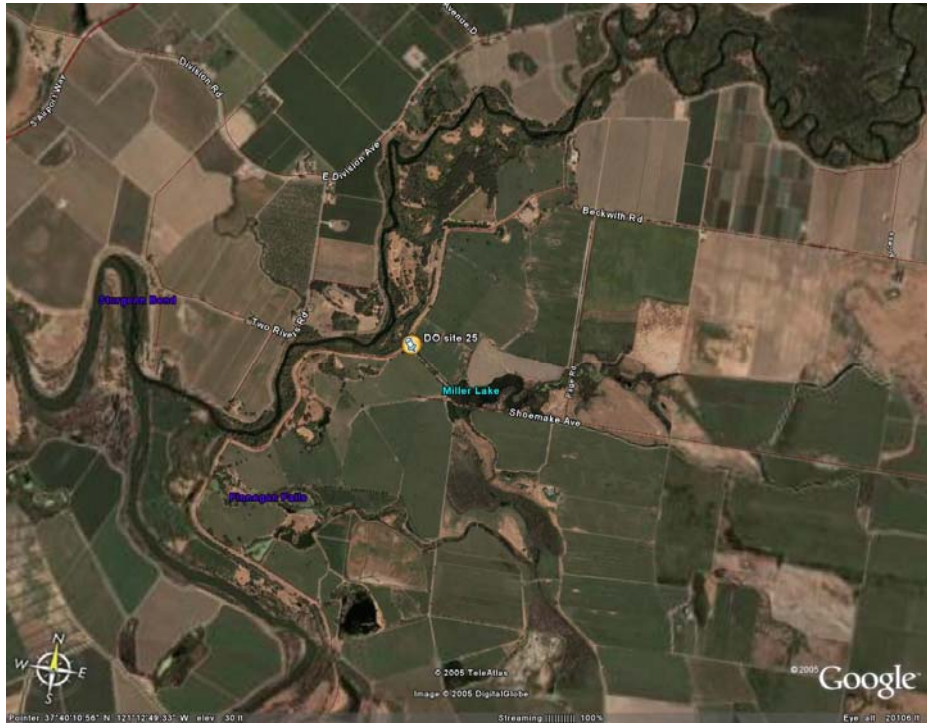


Figure 17: Aerial view of DO TMDL Project water quality sampling station DO-28 Turlock Irrigation District Westport Drain Flow Station.



Figure 18: Aerial view of DO TMDL Project water quality sampling station DO-29 Turlock Irrigation District Harding Drain at Carpenter Road.



Figure 19: Aerial view of DO TMDL Project water quality sampling station DO-36 Del Puerto Creek Flow Station



Figure 20: Aerial view of DO TMDL Project water quality sampling station DO-59 San Joaquin River at Laird Park.

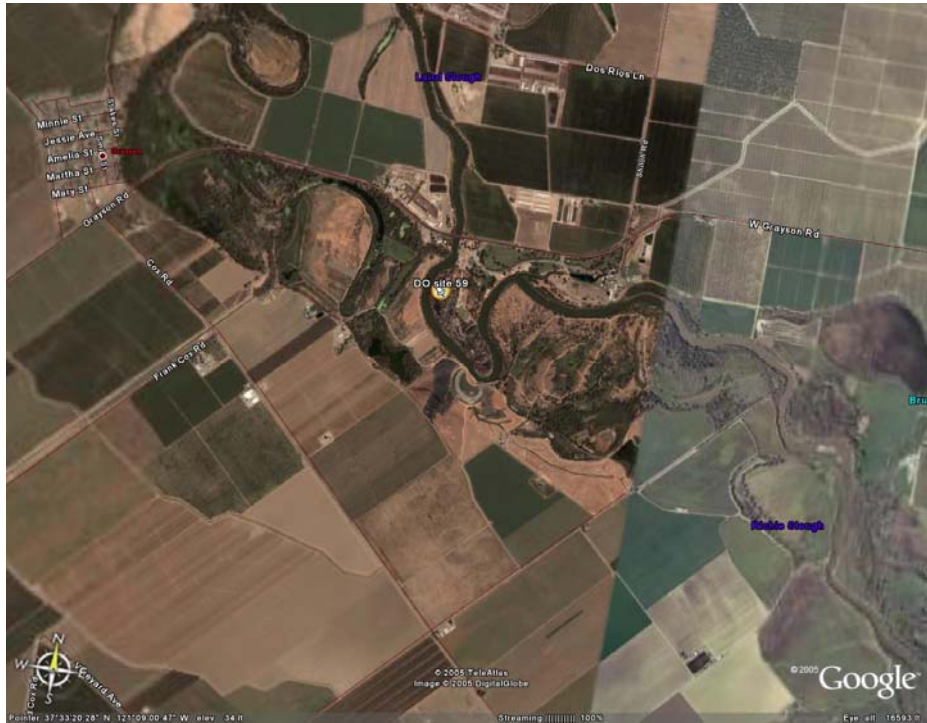


Figure 21: Sampling at DO-5, the McCune (Vernalis) sample station, built and operated by the DWR.



Figure 22: Sampling with bucket at DO-16, Merced River at River Road.



Figure 23: Sample station DO-8, Crows Landing, is located in the Turlock Sportsman Club. In the spring, water levels were to the edge of the upper road shown in picture.



Figure 24: Depth integrated sampling at DO-44, San Luis Drain End. The pole and bottle holder are designed to fill bottles slowly as they decent through the water column.



Figure 25: In-situ measurement of chlorophyll-a and other water quality parameters using a YSI 6600 sonde at DO-21, Orestimba Creek. The tripod allows the sonde to be placed further into the middle of the flow without wading.



Figure 26: University of the Pacific van, equipped for sampling.



Figure 27: Measurement of incident light with spherical and flat quantum light detectors. Light is measured to provide reference for interpretation of chlorophyll-a results.



Figure 28: Example of raw stage data. Stage data will be used to calculate flow data as quality control and calibration data becomes available.

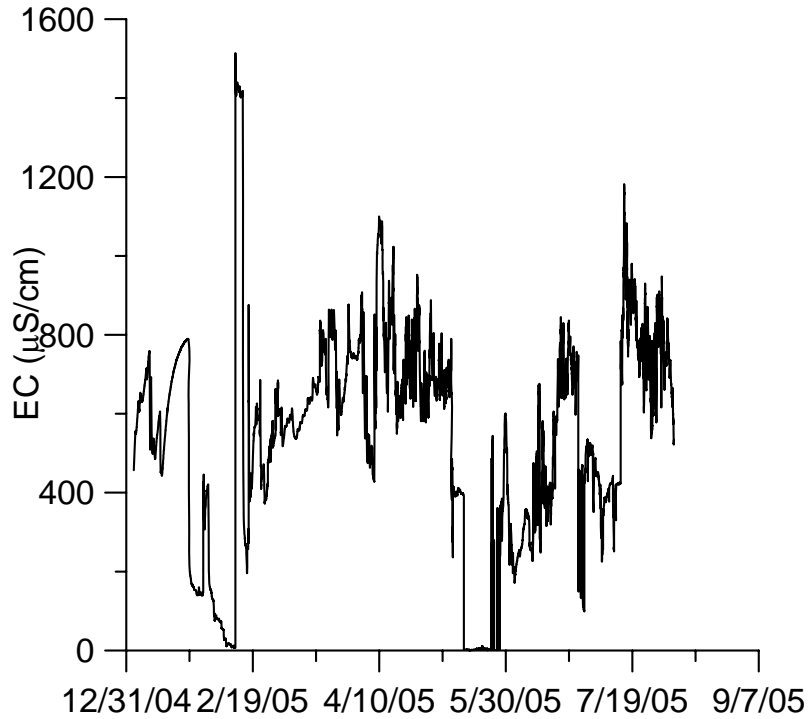


Figure 29: Example of raw electrical conductivity (EC) data being provided by water districts. EC raw data is will be processed into final data when quality control and calibration data becomes available.

